## MS9-02 Time-Resolved Serial Femtosecond Crystallography On Photoswitchable Fluorescent Proteins

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Reversibly photoswitchable fluorescent proteins (RSFP) are essential tools in advanced fluorescence nanoscopy of live cells (such as PALM and RESOLFT). They can be repeatedly toggled back and forth between a fluorescent (on) and a non-fluorescent (off) state by irradiation with light at two different wavelengths. Mechanistic details of photoswitching, in particular on the ultra-fast photochemical time scale, remain largely unknown. Our consortium combines time-resolved serial femtosecond crystallography (TR-SFX) at X-ray free electron lasers, ultrafast absorption spectroscopy in solution and intermediates in two RSFP on the picosecond to microsecond time scale. Two major bottlenecks had to be passed before TR-SFX could be conducted, i.e. the production of well-diffracting microcrystals in large quantities and efficient inline pre-illumination to photoswitch RSFP microcrystals from the on to the off state prior to injection. First pump-probe TR-SFX experiments were conducted at both the LCLS and SACLA that, together with time-resolved absorption spectroscopy, provide first insight into a possible sequence of events involved in photoswitching.

**Keywords:** time-resolved serial femtosecond crystallography, XFEL, microcrystals, fluorescent proteins, chromophore

## MS9-O3 Changes in metal coordination are required to regulate activity of bacterial phosphodiesterases, implicated in c-di-GMP regulated biofilm dispersal

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Many chronic infections are underpinned by the ability of bacteria to transition to the biofilm life-style, which is up to 1000 fold more tolerant to antibiotics. Both biofilm formation and dispersal are controlled by the secondary guanosine bis-(3'-5') cyclic dimeric messenger monophosphate (c-di-GMP); high levels of c-di-GMP are associated with biofilm formation, while a reduction in c-di-GMP induces biofilm dispersal<sup>(1)</sup>. Future pharmaceutical strategies may interfere with these mechanisms. We look at biofilm dispersal, studying c-di-GMP hydrolysis by bacterial phosphodiesterases of the EAL type.

Structures of the EAL type enzymes are readily observed in the substrate bound state (c-di-GMP). These structures pose the question of full enzyme activation; while dimerisation is known to play a key role, many of the structures are observed as dimers<sup>(2,3)</sup>. We present an analysis of these dimers and show that dimerisation alone is insufficient for activation.

Our study of the *Pseudomonas aeruginosa* protein MorA demonstrates that dimerisation is linked with reorganisation of the catalytic site, by unwinding of a helical segment (figure 1)<sup>(4)</sup>. However, this structure is again observed in the substrate bound state. We now present further evidence from phosphodiesterase structures in complex with the hydrolysis product 5'-phosphoguanylyl-(3'-5')-guanosine. Observed differences in metal coordination in the catalytic centre may represent the final layer of enzyme activation. Understanding the full catalytic potential of EAL-type phosphodiesterases is required to explore this class of enzymes in drug design.

1. Schirmer T, Jenal U. "Structural and mechanistic determinants of c-di-GMP signalling." Nat Rev Microbiol. 2009; 7 (10): 724–35.

 Sundriyal A, Massa C, Samoray D, Zehender F, Sharpe T, Jenal U, Schirmer T. "Inherent regulation of EAL domain-catalyzed hydrolysis of second messenger cyclic di-GMP." J Biol Chem. 2014; 289 (10): 6978–90.

3. Winkler A, Udvarhelyi A, Hartmann E, Reinstein J, Menzel A, Shoeman RL, Schlichting I. "Characterization of elements involved in allosteric light regulation of phosphodiesterase activity by comparison of different functional BlrP1 states." J Mol Biol. 2014; 426 (4): 853–68.

4. Phippen CW, Mikolajek H, Schlaefli HG, Keevil CW, Webb JS, Tews I. "Formation and dimerization of