

**P.04.26.16***Acta Cryst.* (2005). **A61**, C269**The Structure of an Ester Synthesising Peptidase**Matthew Bennett<sup>a</sup>, Bryan Anderson<sup>a</sup>, Ross Holland<sup>b</sup>, Gillian Norris<sup>a</sup>,  
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X-prolyl dipeptidyl peptidase (PepX) is a dipeptidase that appears to be ubiquitous in dairy lactic acid bacteria. PepX is best characterised for its highly specific peptidase activity, namely the ability to remove dipeptides from the N-terminus of larger peptides, where proline is residue 2 in the peptide sequence. PepX however is also able to synthesise esters via a transferase mechanism.

The structure of PepX from *Streptococcus thermophilus* has been solved by molecular replacement methods to 1.9 Å resolution using PepX from *Lactococcus lactis* [1] as a model. The refined structure has an R factor of 18.2% and R<sub>free</sub> of 23%.

Characterisation of the ester synthetic activity showed that PepX was capable of producing ethyl butanoate, only if the synthetic triacylglyceride tributyrin was the donor molecule. The basis for this specificity is discussed in terms of the structure of the enzyme, and the topology of the active site. A model for the catalytic activity that is in agreement with the observed kinetic data is presented.

[1] Rigolet P., Mechin I., Delage M.M., Chich J.F., *Structure*, 2002, **10**, 1383.**Keywords:** peptidase, transferase, enzyme kinetics**P.04.26.17***Acta Cryst.* (2005). **A61**, C269**Protein Crystallography with Spallation Neutrons**Benno P. Schoenborn, Paul Langan, *Bioscience Division, Los Alamos National Laboratory, University of California, Los Alamos, NM 87545 USA.* E-mail: schoenborn@lanl.gov

The Protein Crystallography Station (PCS) at the Los Alamos Neutron Scattering Center, is a high performance neutron protein crystallography beam line.[1] Beam time is free and is allocated by a peer review process.

The beam line exploits the pulsed nature of spallation neutrons with a large position sensitive electronic neutron detector that allows time resolved collection of Laue patterns. The data collected uses neutrons with wavelength of 0.7 to 6 Angstroms. The neutron optics employs a partially decoupled moderator with a conical beam line, collection all useful neutrons from the whole moderator surface with a beam divergence matched to the average mosaic of typical protein crystals. [2]

Some typical results from the user program will be presented illustrating data collected for protein crystals with molecular weights from a few kD to over 500kD.

[1] Langan P., Greene G., Schoenborn B.P., *J. App. Cryst.*, 2004, **37**, 24. [2] Benno P. Schoenborn, John D. Court, Allen C. Larson, Phil Ferguson, *J. of Neutron Research*, 1999, **7**, 89-106.**Keywords:** protein crystallography, neutron diffraction, time resolved**P.04.26.18***Acta Cryst.* (2005). **A61**, C269**Crystal Structure of *Morone saxatilis* F-lectin**Mario A. Bianchet<sup>1</sup>, Eric W. Odom<sup>2</sup>, Gerardo R. Vasta<sup>2</sup>, L. Mario Amzel<sup>1</sup>,  
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F-lectins are a novel carbohydrate recognition domain first identified in *Anguilla anguilla* agglutininin (AAA), a 17 kDa serum fucoslectin from European eel [1]. Because AAA specifically recognizes fucosylated oligosaccharides has been used extensively as a reagent in blood typing and histochemistry. F-lectins from invertebrate and vertebrate species function as innate immunity recognition molecules.

The serum of *M. saxatilis*, striped bass, contains a fucoslectin (MsFBP32) that displays two distinct F-lectin sequences in tandem. The crystal structure of MSFB32 complexes with fucose and Lewis-a trisaccharide were determined. The MsFBP32 crystal structure shows a 83 Å long trimer with each distinct monomer CRD segregated to opposite sides. Despite trimers are not observed in solution, each half of the crystal asymmetric unit present striking similarities with the AAA physiological trimer. This arrangement of CRDs suggests a specific function for the recognition of carbohydrates structures on the cellular wall of fish pathogens. Although the two carbohydrate recognition sites of MsFBP32 are F-type carbohydrate binding sites, differences between them suggest that this is a divalent lectin that may recognize and link self to non-self carbohydrate structures.

[1] Bianchet M.A., Odom E.W., Vasta G.R., Amzel L.M., *Nature & Structural Biology*, 2002, **9**, 628-634.**Keywords:** lectin, innate immunity, crystal structure**P.04.26.19***Acta Cryst.* (2005). **A61**, C269**Producing Diffraction Quality Powders from Soluble Lysozyme and Thaumatin**Marc Allaire<sup>a</sup>, Natalia Moiseeva<sup>a</sup>, Peter Stephens<sup>b</sup>, Cristian Botez<sup>b</sup>,  
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The pioneer effort on insulin and lysozyme has revealed the possibility of acquiring powder diffraction profile from proteins. These powder profiles were shown to be of sufficient quality to extract structural information. These results imply the idea of using protein powder diffraction for the identification of ligand complexes.

Our effort is to develop a general method to obtain polycrystalline powder from protein in solution. Our approach takes advantage of the crystallization conditions known to produce single crystal. Lysozyme and thaumatin were used as test case in this study. The crystallization conditions explored for lysozyme were from NaCl in Acetate buffer pH 4.5 and the Na/K tartrate in MES buffer pH 6.5 for thaumatin.

In order to generate protein powder, we increased the number of nucleation sites by increasing the concentration of protein and/or precipitant. In both cases, the proteins were first dissolved in the appropriate buffer and then the precipitant was added. Powder diffraction profiles were collected on the high-resolution powder beam line X3B1 at the National Synchrotron Light Source and could be interpreted from the known single crystal lattice. Our results suggest that polycrystalline powder can be produce from soluble lysozyme and thaumatin and further analysis is in progress to apply this approach to other proteins.

**Keywords:** protein crystallography, powder diffraction, protein crystallization**P.04.26.20***Acta Cryst.* (2005). **A61**, C269-C270**A Novel Chlorophyll-binding Mode of Water-soluble Chlorophyll Protein (WSCP)**Daisuke Horigome<sup>a,b</sup>, Hiroshi Hara<sup>b</sup>, Hiroyuki Satoh<sup>b</sup>, Atsushi Nakagawa<sup>a</sup>, Akira Uchida<sup>b</sup>,  
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Chlorophyll (Chl), the most important pigment in photosynthesis, is known as a generator of oxygen radical under excess light. Since the oxygen radical is harmful on plant cellular component, plants need to quench it. In the photosynthetic apparatus, carotenoid quenches the overexcited Chl by xanthophyll cycle. However, it still remains to be seen that how plants prevent the Chl-mediated oxygen radical formation at the stage of Chl biosynthesis and Chl transport pathway.

The putative Chl carrier, water-soluble chlorophyll protein (WSCP), prevents Chl-mediated oxygen radical formation without carotenoid in an as yet unknown manner [1]. To elucidate this mechanism, we crystallized the WSCP from *Brassica oleracea* var.