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Crystal structure of the pyoverdinin outer membrane receptor FpvA from *Pseudomonas aeruginosa*. Cobessi D., Célia H., Folschweiler N., Schalk I., Abdallah M. & Pattus F. *Département Récepteurs et Protéines Membranaires, UPR9050 CNRS, Ecole Supérieure de Biotechnologie de Strasbourg, Boulevard Sébastien Brant, 67410 Illkirch, France. Email: cobessi@esbs.u-strasbg.fr*

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When grown under iron-deficient conditions, many bacteria synthesize and release into the environment iron chelators termed siderophores. In the host it is expected that siderophores sequester iron from iron-containing molecules such as transferrin and deliver iron to the microbial cell. In general, the first step of entry of ferric siderophores into Gram negative bacteria is mediated by specific outer membrane receptors. 3 x-ray structures of the outer membrane ferric siderophore receptors from *E. coli* have been reported. This transport into the periplasm requires the cytoplasmic proton motive force and an energy transduction complex which includes the cytoplasmic membrane proteins TonB, ExbB and ExbD [1]. *P. aeruginosa* is an opportunistic human pathogen which infects injured, immunodeficient, or otherwise compromised patients. Under iron-limited conditions, the bacterium secretes a major siderophore called pyoverdinin (PaA). PaA seems to play an important role in infection by competing with transferrin for iron in order to overcome the iron-withholding mechanism present in mammals. Previous *in vitro* and *in vivo* studies have shown that pyoverdinin mediated iron uptake through its outer membrane receptor FpvA occurs through a novel mechanism different from the uptake mediated by ferrichrome in *E. coli*. FpvA is able to bind both iron free PaA and ferric-PaA and the normal state of the FpvA receptor under iron limitation seems to be the FpvA-PaA complex. During iron uptake, the extracellular ferric-PaA displaces the bound PaA on the FpvA receptor [2]. We over-expressed and purified FpvA (MM: 86245) from *P. aeruginosa* in its siderophore-bound (FpvA-PaA) and in its ligand-free (FpvA) forms. We crystallized the different forms of the receptor. We are currently building a first atomic model of FpvA-PaA at 3.6 Å resolution from a dataset collected using crystals of a SeMet substituted FpvA-PaA. The current free-R and R factors are now below 30 %. Three molecules related by a non crystallographic 3-fold axis are in the asymmetric unit. The FpvA-PaA model contains 684 residues and the pyoverdinin. The structure can be divided in two domains. The N-terminal part of the structure called cork-domain contains 146 residues. It fills the second domain composed of 538 residues which is a β -barrel of 22 antiparallel transmembrane strands. The siderophore is bound on the extracellular side of the receptor where the loops connecting the β -strands are larger than the ones situated in the periplasm. This structure is the first of an *in vivo* loaded siderophore receptor and also the first TonB-receptor structure from another bacterium than *E. coli*.

[1] Braun, V. and Braun, M. (2002). *FEBS Lett.*, **529**, 78-85.

[2] Schalk, I.J., Abdallah, M.A. and Pattus, F. (2002). *Biochem. Soc. Trans.*, **30**, 702-705.

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Crystal structure of mouse CD1d with a phospholipid self ligand: insights in the molecular basis of regulatory NKT cell activation. Massimo Degano^a, Barbara Giabbai^a, Stéphane Sidobre^b, Yovan Sanchez-Ruiz^c, Angela Bachi^c, Mitchell Kronenberg^b. ^a*Biocrystallography Unit and cMass Spectrometry Unit, DIBIT San Raffaele Scientific Institute, I-20132 Milan, Italy,* ^b*Division of Developmental Immunology, La Jolla Institute for Allergy and Immunology, 92121 La Jolla CA, USA. E-mail: degano.massimo@hsr.it*

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NKT cells are immunoregulatory lymphocytes able to modulate the immune response through rapid secretion of cytokines. These cells are known to promote spontaneous rejection of tumors, or control an unwanted immune reaction in several autoimmune pathologies. NKT cells are activated is by the recognition of glycolipid antigens bound to the CD1d molecules by a semi-invariant T cell antigen receptor (TCR). The molecular details of the interactions between CD1d and its ligands, however, are still elusive. Here we present the crystal structure to 2.8Å of mouse CD1d bound to the self ligand phosphatidyl choline. The structure of the CD1d/ligand complex allows the definition of the structural and chemical requirements for the binding of lipids to group II CD1 molecules. The shape and volume of the binding groove limit the pool of CD1d-restricted lipid antigens to compounds with defined acyl chain composition. The orientation of the antigen polar headgroup towards the C-terminus of the α 1 helix provides a rationale for the structural basis for the observed V α chain bias in NKT cells. The contribution of the ligand to the protein surface suggests a likely mode of recognition of lipid antigens by the NKT cell TCR that is distinct from the classic TCR/MHC/peptide interaction. The structure of the CD1d/self antigen complex suggests rational modifications of known antigens that could modulate the activity *in vivo* of NKT cells.