

03-Crystallography of Biological Macromolecules

119

PS-03.11.10

PRELIMINARY CRYSTALLOGRAPHIC STUDIES OF GROTIIN II. By Chen Minghuang, Zhou Kangjing, Fu Zhuji and Pan Kezhen*, Fujian Institute of Research on the Structure of Matter, Chinese Academy of Sciences, Fuzhou 350002, and National Lab. of Biomacromolecule, Beijing 100101.

Two plant toxins, crotin I and crotin II have been isolated and purified from the seeds of Euphorbiaceae *Croton tiglium*, which is a Chinese medicinal herb named Ba Dou. The molecular weight (M.W.) of crotin I and crotin II measured by SDS-PAGE are about 40,000 and 15,000 Da, respectively. (Chen M.H. & Pan K.Z., Chinese Biochemical J., in press). It was observed that these two proteins inhibit protein synthesis in a cell-free system (Stirpes, F. et al, 1976, Biochem. J. 156, 1-6) and depurinated rat liver ribosomes (Barbieri, L. et al., 1992, Biochem. J. 286, 1-4). They belong to so-called ribosome inactivating proteins (RIPs). The experiments show that crotin II has much higher inhibitory activity than crotin I. Crotin II has a neutral pI and a lower M.W., it is different from single chain RIPs (e.g. Trichosanthin), which have a basic pI and a higher M.W. (e.g. 27,000 Da). However, they have a similar function to inhibit protein synthesis. So the study of the three-dimensional structure of crotin II is important in the relationship of structure and function in single chain RIPs.

The crystallization was performed by using the hanging-drop method. The crystals of crotin II with high quality were grown at room temperature in a citrate buffer solution with KCl as the precipitant. The crystals grow to a size of 0.7 mm x 0.3 mm x 0.3 mm within ten days. Precession photographs of the crystals mounted in the thin-wall siliconized glass capillary tubes were taken by using a Ni-filtered CuK α radiation (40 KV, 100mA). The cell parameters were determined to be $a = b = 94.62 \text{ \AA}$, $c = 28.43 \text{ \AA}$, $\alpha = \beta = 90^\circ$, $\gamma = 120^\circ$. The extinction rules and intensity distribution of the reflections show that the crystal belongs to space group P6₃ or P6₅. Assuming one molecule in an asymmetric unit, the V_m value of 2.46 $\text{\AA}^3/\text{Da}$ and 44% solvent contents were calculated (Matthews, B.M., 1968, J.Mol. Biol., 33, 491).

X-ray diffraction data for native crystals were collected on area detector (Siemens X-200B). Each oscillation frame covered 0.25° and was measured for 120s. Total 720 frames were collected. The data were reduced by using the XGEN program. Final merged diffraction data have 11,822 unique reflections within the 1.82 \AA resolution. $R_{\text{merge}} = 0.0819$.

PS-03.11.11 STRUCTURE OF ORTHORHOMBIC CRYSTAL OF TRICHOSANTHIN AT 1.88 \AA RESOLUTION. By Zhou Kang-jing*, Fu Zhu-ji, Chen Ming-huang, Lin Yu-juan and Pan Ke-zhen, Fujian Institute of Research on the Structure of Matter, Chinese Academy of Sciences, Fuzhou, 350002, Fujian, China.

Trichosanthin is one of the ribosome inactivating proteins (RIPs) extracted from a Chinese herb medicine, the root tuber of *Trichosanthes kirilowii*, Maxim Cucurbitaceae. It consists of 247 amino acids with $M_r = 27,132$. The orthorhombic crystals of trichosanthin have been obtained by using hanging drop method under the condition of pH=5.6. The crystal belongs to the space group P2₁2₁2₁ with $a = 38.305$, $b = 76.221$, $c = 79.213 \text{ \AA}$. The X-ray intensity data of 15466 reflections were collected on a Siemens X-200B area detector. The structure was solved by molecule replacement methods using the model of trichosanthin molecule of monoclinic crystal as the known

structural model. The initial model was refined using the programs of XPLOR and PROLSQ to an R-factor of 0.191 for the reflections between 6 \AA -1.88 \AA . The r.m.s. deviations of bond length and bond angle are 0.013 \AA and 0.055°, respectively. Trichosanthin molecule can be divided into two structural domains with different size. The molecule contains 8 α helices and 13 β strands, the characteristic of which is that almost all α helices were in the inner of the molecule, whereas all β strands were near the surface. The active site of the molecule consisted of 5 conservative residues is located on the concave region between the two domains. In the active site Arg122 and Glu189, Arg163 and Glu160 form two ion pairs, Glu189 and Gln156 are hydrogen bonded to each other. A total of 219 solvent molecules are included in the final refined model. Comparing with the structure of monoclinic crystal of trichosanthin grown under the condition of pH=8.4, it is shown that there are little differences between the two structures.

PS-03.11.12

THE REFINED CRYSTAL STRUCTURE OF THE NEUROPHYSIN-OXYTOCIN COMPLEX AT 2.8 \AA RESOLUTION. John P. Rose* and Bi-Cheng Wang, Departments of Crystallography and Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260 USA.

The posterior pituitary produces two important regulatory hormones, oxytocin and vasopressin. Oxytocin is known to mediate uterine contraction and milk ejection and has recently been shown to play an important role in sexual behavior and response, as well as bonding between parent and offspring. Vasopressin plays an important role in influencing kidney function, blood pressure and body fluids. Both hormones are nonapeptides and are found in concentrations as high as 0.1 M in the neurosecretory granules of the posterior pituitary complexed in a 1:1 ratio with a class of small (11 kD) disulfide-rich proteins called neurophysins.

Single crystals of a bovine neurophysin II - oxytocin complex have been obtained using (NH₄)₂SO₄ as the precipitating agent (Rose *et al.* (1991) *J. Mol. Biol.* 221, 43). The crystals diffract to at least 3 \AA resolution, belong to Laue group 4/mmm and exhibit systematic absences consistent with either space group P4₁2₁2 or P4₃2₁2. The cell dimensions are $a = b = 69.07 \text{ \AA}$ and $c = 113.26 \text{ \AA}$. The crystals contain one neurophysin-oxytocin dimer per asymmetric unit. Based on a V_m of 2.9 $\text{\AA}^3/\text{dalton}$, the solvent content is calculated to be 58%. The structure of the protein-hormone complex has been determined by molecular replacement using the structure of a bovine neurophysin II Phe-Tyr-NH₂ complex (Chen, *et al.* (1991) *Proc. Natl. Acad. Sci. USA*, 88, 4240) as the search model. A full crystallographic refinement of the neurophysin-oxytocin complex is underway. Details of the structure and crystallographic analysis will be presented.

Work supported by NIH grant GM-46828 and a grant from the Pittsburgh Supercomputing Center.

PS-03.11.13 THE REFINED CRYSTAL STRUCTURE OF A NEUROPHYSIN-DIPEPTIDE COMPLEX AT 2.5 \AA RESOLUTION. Chia-Kuei Wu*, John P. Rose and Bi-Cheng Wang, Departments of Crystallography and Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260 USA.

Neurophysins are small disulfide rich proteins. They are found in concentrations as high as 0.1 M in the neurosecretory granules of the posterior pituitary where they are involved in the binding and storage of the posterior pituitary hormones oxytocin and vasopressin.

The crystal structure of a complex of NP-II (a vasopressin-associated NP) with I-Phe-Tyr-NH₂ which binds at the hormone-binding site was determined by using single wavelength anomalous scattering data (Chen, *et al.* (1991) *Proc. Natl. Acad. Sci. USA*, 88, 4240). It is an intermediate step in solving the structure of the native neurophysin - dipeptide complex.

The native NP-II-Phe-Tyr-NH₂ complex was crystallized in space group P2₁2₁2₁ and diffracts to 2.5 \AA . In this structure, which is